

### **DNA-PAINT KIT**

## **MASSIVE-sdAB-FAST 2-PLEX**

Expiration after 6 months

(For research use only)

#### **CONTENT**

#### **SECONDARY SINGLE DOMAIN ANTIBODIES**

- FluoTag®-XM-QC Anti-Mouse IgG kappa light chain (Clone: 1A23) + FAST docking site F1 (To be measured with Imager F1)
- FluoTag®-XM-QC Anti-Rabbit IgG (Clone: 10E10) + FAST docking site F2 (To be measured with Imager F2)
- Concentration: 5 μM Protein, 5 μM DNA (1 DNA strand per protein)
- Volume: 100 μL
- Storage: -20 °C
- Storage buffer: PBS, 50 % glycerol + 0.05 % NaN<sub>3</sub>
- Recommend dilution: 1:100 1:500 (for optimal results the dilution needs to be optimized depending on the target accessibility and expression level)

#### **IMAGERS**

- Imager F1 Cy3B or ATTO 655
- Imager F2 Cy3B or ATTO 655
- Concentration: 1 μM in TE buffer (10 mM Tris, 1 mM EDTA, pH 8)
- Volume: 300 µL
- Storage: 20 °C (1 μM imager solutions are stable for multiple freeze-and-thaw cycles)

Optional: Prepare 50  $\mu$ L aliquots and store at -20 °C. Working aliquots can be stored at 4 °C for short-term or -20 °C for long term

Note: Further dilutions should be prepared fresh before use. Low imager concentrations are not stable in plastic tubes.

#### **BUFFERS**

Antibody incubation buffer, 30 mL, store at 2-8 °C

Note: For longer-term storage we recommend to store aliquots at -20  $^{\circ}\text{C}.$ 

- Washing buffer (10×), 20 mL, store at room temperature (to be diluted 1:10 in water before use)
- Imaging buffer, 50 mL, store at room temperature

# SAMPLE PREP. PROTOCOL

- 1. Prepare sample using a protocol optimized for your target and primary antibody staining.
- 2. Wash with washing buffer (1×).
- 3. Dilute secondary DNA-PAINT single domain antibodies in Antibody incubation buffer.
- 4. Incubate for 30 60 min at room temperature.
- 5. Wash three times with washing buffer (1×).
- 6. Optional: Incubate fiducial markers.
- 7. Wash with imaging buffer before adding the final imaging solution with imager strands.
- 8. Before imaging: Add imager strands diluted in imaging buffer. We recommend a starting concentration of 1 nM. However, the optimum imager concentration strongly depends on the target and labeling density. Thus, the imager concentration should be adjusted such that distinct single molecule blinking events can be observed.
- 9. After imaging, exchange buffer to washing buffer (1x) for storage at 4  $^{\circ}$ C.

## IMAGING PARAMETERS

- Exposure time: 30 50 ms
- Laser-Intensity: ~300 W/cm² (561 nm) and ~500 W/cm² (640 nm). This intensity might vary due to different
  illumination depths/modes (TIRF/HILO). For dense targets we recommend increasing the laser power to enhance
  blinking.
- Total imaging time/target: 5-15 min (Depends on target density and applied imager concentration)
- Temperature: The kit is optimized for image acquisition at 21 25 °C. At higher temperatures shorter exposure times and higher laser powers are required.



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