

DNA-PAINT KIT

MASSIVE-sdAB-FAST 2-PLEX

Expiration after 6 months

(For research use only)

CONTENT

SECONDARY SINGLE DOMAIN ANTIBODIES

- **FluoTag®-XM-QC Anti-Mouse IgG kappa light chain** (Clone: 1A23) + **FAST docking site F1** (To be measured with Imager F1)
- **FluoTag®-XM-QC Anti-Rabbit IgG** (Clone: 10E10) + **FAST docking site F2** (To be measured with Imager F2)
- Concentration: 5 μM Protein, 5 μM DNA (1 DNA strand per protein)
- Volume: 100 μL
- Storage: $-20\text{ }^{\circ}\text{C}$
- Storage buffer: PBS, 50 % glycerol + 0.05 % NaN₃
- Recommended dilution: 1:100 - 1:500 (for optimal results the dilution needs to be optimized depending on the target accessibility and expression level)

IMAGERS

- **Imager F1** Cy3B or ATTO 655
- **Imager F2** Cy3B or ATTO 655
- Concentration: 1 μM in TE buffer (10 mM Tris, 1 mM EDTA, pH 8)
- Volume: 300 μL
- Storage: $-20\text{ }^{\circ}\text{C}$ (1 μM imager solutions are stable for multiple freeze-and-thaw cycles)
Optional: Prepare 50 μL aliquots and store at $-20\text{ }^{\circ}\text{C}$. Working aliquots can be stored at $4\text{ }^{\circ}\text{C}$ for short-term or $-20\text{ }^{\circ}\text{C}$ for long term
Note: Further dilutions should be prepared fresh before use. Low imager concentrations are not stable in plastic tubes.

BUFFERS

- **Antibody incubation buffer**, 30 mL, store at $2\text{--}8\text{ }^{\circ}\text{C}$
Note: For longer-term storage we recommend to store aliquots at $-20\text{ }^{\circ}\text{C}$.
- **Washing buffer (10 \times)**, 20 mL, store at room temperature (to be diluted 1:10 in water before use)
- **Imaging buffer**, 50 mL, store at room temperature

SAMPLE PREP. PROTOCOL

1. Prepare sample using a protocol optimized for your target and primary antibody staining.
2. Wash with washing buffer (1 \times).
3. Dilute secondary DNA-PAINT single domain antibodies in Antibody incubation buffer.
4. Incubate for 30 - 60 min at room temperature.
5. Wash three times with washing buffer (1 \times).
6. Optional: Incubate fiducial markers.
7. Wash with imaging buffer before adding the final imaging solution with imager strands.
8. Before imaging: Add imager strands diluted in imaging buffer. We recommend a starting concentration of 1 nM. However, the optimum imager concentration strongly depends on the target and labeling density. Thus, the imager concentration should be adjusted such that distinct single molecule blinking events can be observed.
9. After imaging, exchange buffer to washing buffer (1 \times) for storage at $4\text{ }^{\circ}\text{C}$.

IMAGING PARAMETERS

- Exposure time: 30 - 50 ms
- Laser-Intensity: $\sim 300\text{ W/cm}^2$ (561 nm) and $\sim 500\text{ W/cm}^2$ (640 nm). This intensity might vary due to different illumination depths/modes (TIRF/HILO). For dense targets we recommend increasing the laser power to enhance blinking.
- Total imaging time/target: 5-15 min (Depends on target density and applied imager concentration)
- Temperature: The kit is optimized for image acquisition at $21\text{--}25\text{ }^{\circ}\text{C}$. At higher temperatures shorter exposure times and higher laser powers are required.

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